# NOR association in Canis familiaris

M Rønne, BS Poulsen, Y Shibasaki

Odense University, Institute of Medical Biology, Department of Anatomy and Cytology, Campusvej 55, DK-5230 Odense M, Denmark

(Proceedings of the 9th European Colloquium on Cytogenetics of Domestic Animals; Toulouse-Auzeville, 10-13 July 1990)

nucleolar organizer region (NOR) / RBG-bands / association / dog

#### INTRODUCTION

The domestic dog has 78 chromosomes (Gustavsson, 1964). Because of the large number of chromosomes of almost similar size and morphology, a precise analysis of the karyotype is rather difficult (Howard-Peebles and Pryor, 1980; Manolache *et al*, 1976; Wurster-Hill and Centerwall, 1982). However, a G-banded karyotype of *Canis lupus* (Wayne *et al*, 1987), which is supported to be homologue to that of *Canis familiaris*, displayed detailed information about banding patterns and served as a guideline for the numbering system used in the RBG-banded karyotype (Poulsen *et al*, in press) and in the present study. Several authors have reported on the nucleolar organizer regions (NORs) and NOR associations in the karyotype of the domestic dog (Kopp *et al*, 1982a, b; Pathak *et al*, 1982). In this paper, the frequencies of NOR associations and the localization of NORs in the female karyotype of *Canis familiaris* are presented using sequential RBG-banding and the silver (Ag)-NOR staining technique.

### MATERIALS AND METHODS

The donor animals were selected at random from the laboratory *Beagles* at the Institute of Biomedicine, Odense University. Peripheral blood samples from 3 female dogs were cultured and processed for RBG-band induction as previously described (Rønne 1985; Poulsen *et al*, in press). Ag-NOR staining was modified after Howell and Black (1980) and used to stain previously RBG-banded metaphases.

From each donor animal, 10 selected RBG-banded metaphase and prometaphase plates with good spreading and well-defined bands were photographed, registered and karyotyped according to Poulsen *et al* (in press). The selected metaphase and prometaphase plates were subsequently silver-stained to display active NORs and rephotographed after counter-staining with 3% Giemsa solution in Sørensen's phosphate buffer (pH 8.0) for 5 min. Comparison between donor animals showed the same NOR pattern for all 3 animals. Forty RBG-banded metaphase and prometaphase plates selected at random from all 3 donor animals were photographed and registered. These cells were then silver-stained, counter-stained with Giemsa and rephotographed. Metaphase and prometaphase plates with corresponding R-band and NOR-staining were compared. The locations of active NORs and NOR associations were determined.

## RESULTS

A total of 40 randomly selected metaphases were sequentially examined with RBGbanding and Ag-NOR staining. Eight autosomes, 5, 8, 14, 16, 19, 21, 32, 37, carry NORs in the telomeric regions (fig 1). The late-replicating X chromosome also showed an active telomeric NOR (Xq) in 10% of the examined metaphases (fig 2). Twenty-one metaphases displayed NOR association among the autosomes at a range of 1–3 associations per metaphase. The X chromosomes were not involved in any NOR association. As shown in table I, chromosomes 14 (38.46%) and 16 (19.23%) have remarkably high levels of involvement in NOR association. Typical NOR associations are shown in figure 3.



Fig 1. The 8 representative autosomes of the dog carrying active NORs after fluorescence plus Giemsa (FPG)-staining (left side) and subsequent Ag-NOR staining (right side).

## DISCUSSION

Several authors have described NORs in domestic dog (Kopp *et al*, 1982a, b; Pathak *et al*, 1982; Howard-Peebles and Howell, 1983). Since the dog karyotype is not standardized, the reported positions of NORs on dog chromosomes were inconsistent. Howard-Peebles and Howell (1983) published that the maximum number of NORs in the dog karyotype was 8, including the NOR on the Y chromosome. Using high-resolution RBG-banding (Poulsen *et al*, in press), NORs



Fig 2. The late-replicating X chromosomes of the domestic dog showing silver-positive NOR (right side) on the telomere of the q-arm.

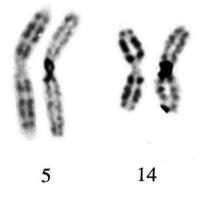


Fig 3. Typical NOR associations observed in dog karyotypes (right side). When stained only with Giemsa (left side), they look like metacentric chromosomes.

Chromosome number	5	8	14	16	19	21	32	37	Total	Frequency
5	1			1				1	3	5.77
8			$^{2}$		1	1	1		5	9.62
14		$^{2}$	$3^{\mathrm{a}}$	$8^{\mathrm{a}}$		3	$^{2}$	<b>2</b>	20	38.46
16	1		$8^{\mathrm{a}}$			1			10	19.23
19		1				1			<b>2</b>	3.85
21		1	3	1	1				6	11.54
32		1	2						3	5.77
37	1		2						3	5.77
Total									52	100.01

Table I. The number and frequency of 26 identified NOR associations between autosomes.

<sup>a</sup> NOR association (14-14-16) was counted as 14-14, 14-16, 14-16.

on 8 different autosomes were observed (fig 1). In males, Pathak *et al* (1982) and Kopp *et al* (1982b) reported that the Y but not the X chromosome displayed active NORs. Late-replicating X chromosomes bearing silver grains were observed at a low frequency (10%) after Ag-NOR staining (fig 2). However, at this stage, further investigation of a larger population is needed to determine the presence and role of NORs on sex chromosomes in the dog.

As previously reported by Kopp *et al* (1982a), we observed a high incidence (50%) of metaphases with NOR association. NOR-associated chromosomes may look like metacentrics as shown in figure 3. Translocations in normal dog (Larsen *et al*, 1978, 1979; Mayr *et al*, 1986; Ma and Gilmore, 1971; Welling and Strandström 1988), dog cancer (Grindem and Buoen, 1986; Benjamin and Noronha, 1967; Oshimura *et al*, 1973; Else *et al*, 1982; Welling *et al*, 1988) and other abnormalities (Shive *et al*, 1965; Hare *et al*, 1967) were reported after using conventional Giemsa-stained or G-banded chromosomes. Especially in cancer studies (Mellink *et al*, 1989), metacentric chromosomes have been used as markers of neoplastic development. Without using NOR staining, however, there is a high risk of confusing NOR association with true translocation.

#### REFERENCES

Benjamin SA, Noronha F (1967) Cytogenetic studies in canine lymphos<br/>arcoma. Cornell Vet 57, 526-542

Else RW, Norval M, Neill WA (1982) The characteristics of a canine mammary carcinoma cell line, REM 134. Br J Cancer 46, 675-681

Grindem CB, Buoen LC (1986) Cytogenetic analysis of leukaemic cells in the dog. A report of 10 cases and a review of the literature. J Comp Pathol 96, 623-635

Gustavsson I (1964) The chromosomes of the dog. Hereditas 51, 187-189

Hare WCD, Wilkinson JS, McFeely RA, Riser WH (1967) Bone chondroplasia and a chromosome abnormality in the same dog. Am J Vet Res 28, 583-587

Howard-Peebles PN, Pryor JC (1980) The R-banding pattern of the canine karyotype. J Hered 71, 361-362

Howard-Peebles PN, Howell WM (1983) Nucleolus organizer regions of the canine karyotype. Cytogenet Cell Genet 35, 293-294

Howell WM, Black DA (1980) Controlled silver-staining of nucleolus organizer regions with a protective colloidal developer: a 1-step method. *Experientia* 36, 1014-1015

Kopp E, Mayr B, Schleger W (1982a) Nucleolus organizer regions on chromosomes of the domestic dog. J Hered 73, 73

Kopp E, Mayr B, Schleger W (1982b) Nucleolus organizer regions on chromosomes of the domestic dog. J Hered 73, 230

Larsen RE, Dias E, Cervenka J (1978) Centric fusion of autosomal chromosomes in a bitch and offspring. Am J Vet Res 39, 861-864

Larsen RE, Dias E, Flores G, Selden JR (1979) Breeding studies reveal segregation of a canine Robertsonian translocation along Mendelian proportions. *Cytogenet Cell Genet* 24, 95-101

Ma NSF, Gilmore CE (1971) Chromosomal abnormality in a phenotypically and clinically normal dog. *Cytogenetics* 10, 254-259

Manolache M, Ross WM, Schmid M (1976) Banding analysis of the somatic chromosomes of the dog (*Canis familiaris*). Can J Genet Cytol 18, 513-518

Mayr B, Krutzler J, Schleger W, Auer H (1986) A new type of Robertsonian translocation in the domestic dog. J Hered 77, 127

Mellink CHM, Bosma AA, Rutteman GR (1989) Chromosome abnormalities in a case of canine metastatic mammary carcinoma. *Proceedings of the 8th European Colloquium on Cytogenetics of Domestic Animals Bristol* (Long S, ed) 115-121

Oshimura M, Sasaki M, Makino S (1973) Chromosomal banding patterns in primary and transplanted venereal tumors of the dog. J Natl Cancer Inst 51, 1197-1203

Pathak S, van Tuinen P, Merry DE (1982) Heterochromatin, synaptonemal complex, and NOR activity in the somatic and germ cells of a male domestic dog, *Canis familiaris* (Mammalia, Canidae). *Cytogenet Cell Genet* 34, 112-118

Poulsen BS, Shibasaki Y, Rønne M (1991) Banding studies in *Canis familiaris*. I. Replication patterns in karyotypes from lymphocyte cultures. *Cytobios*, in press

Rønne M (1985) Double synchronization of human lymphocyte cultures: selection for high resolution banded metaphases in the first and second division. Cytogenet Cell Genet 39, 292-295

Shive RJ, Hare WCD, Patterson DF (1965) Chromosome studies in dogs with congenital cardiac defects. *Cytogenetics* 4, 340-348

Wayne RK, Nash WG, O'Brien SJ (1987) Chromosome evolution of the Canidae. I. Species with high diploid numbers. Cytogenet Cell Genet 44, 123-133

Welling J, Strandström H (1988) Unbalanced Robertsonian translocations in canine lymphocytes and bone marrow cells *in vitro*. *Hereditas* 109, 193-196

Welling J, Strandström H, Knuutila S (1988) Lymphatic leukaemia cell line 3447 from the dog. A karyotypic analysis. *Hereditas* 109, 185-191

Wurster-Hill DH, Centerwall WR (1982) The interrelationships of chromosome banding pattern in canids, musterids, hyena, and felids. Cytogenet Cell Genet 34, 178-192