SHORT COMMUNICATION







Application of mixed linear models for the estimation of functional effects on bovine stature based on SNP summary statistics from a whole-genome association study

Krzysztof Kotlarz¹, Barbara Kosinska-Selbi¹, Zexi Cai², Goutam Sahana² and Joanna Szyda^{1,3*}

Abstract

Genome-wide association studies (GWAS) help identify polymorphic sites or genes linked to phenotypic variance, but a few identified genes and/or single nucleotide polymorphisms (SNPs) are unlikely to explain a large part of the phenotypic variability of complex traits. In this study, the focus was moved from single loci to functional units, expressed by the metabolic pathways as defined in the Kyoto Encyclopaedia of Genes and Genomes (KEGG) database. Consequently, the aim of this study was to estimate KEGG effects on stature in three Nordic dairy cattle breeds using SNP effects from GWAS as the dependent variable. The SNPs were annotated to genes, then the genes to KEGG pathways. The effects of KEGG pathways were estimated separately for each breed using a mixed linear model incorporating the similarity between pathways expressed by common genes. The KEGG pathway D-amino acid metabolism (map00473) was estimated to be significant for stature in two of the analysed breeds and revealed a borderline significance in the third breed. Thus, we demonstrate that the approach to statistical modelling of higher order functional effects on complex traits is useful, and provides evidence of the importance of D-amino acids for growth in cattle.

Background

Genome-wide association studies (GWAS) are very useful for the identification of polymorphic sites, typically single nucleotide polymorphisms (SNPs), or sometimes of genes associated with a phenotypic variation or with a disease. Today, the common availability of SNPs obtained based on whole-genome sequencing allows for a very good resolution of the estimation of those associations. However, in the context of phenotypes that show a complex mode of inheritance, a few genes and/or SNPs are not expected

*Correspondence: joanna.szyda@upwr.edu.pl

to be sufficient to explain the variability on a phenotypic level. As a consequence, in many cases, it is possible to identify loci with a very large effect on the phenotypic variation, but still, a predominant proportion of this variation remains unexplained [1], since it is often due to a combined effect of many loci, each with a moderate or small impact. Therefore, in our study, we moved the focus from a single locus to functional units, here expressed by the metabolic pathways defined by the Kyoto Encyclopaedia of Genes and Genomes (KEGG) database. This approach allows us to better understand the physiological mechanisms that underlie complex phenotypes. In the literature, the estimation of the effects of pathways has mainly been applied to gene expression data, but rarely in the GWAS context (see e.g. [2]). In this study, we used SNP summary statistics



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

¹ Biostatistics Group, Department of Genetics, The Wroclaw University of Environmental and Life Sciences, Kozuchowska 7, 51-631 Wroclaw, Poland Full list of author information is available at the end of the article

originating from the GWAS conducted for stature and based on whole-genome sequence data of three Nordic dairy cattle breeds. To our knowledge, this methodological approach proposes a new model, that is statistically most similar, albeit not equivalent, to the analysis described by [3].

Methods

The analysed data comprised SNP summary statistics from GWAS performed on 5062 Danish Holstein bulls, 924 Danish Red Dairy Cattle bulls, and 2122 Finnish Red Dairy Cattle bulls [4]. The association was calculated for 25.4 million variants that were imputed with Minimac2 [5] from 630,000 SNPs using the 1000 Bull Genomes reference population from Run4, consisting of 1147 individuals. SNP additive effects were estimated for deregressed estimated breeding values (EBV) for stature. These were used as pseudophenotypes, separately for each breed, with a single SNP mixed linear model including an additive polygenic effect with a covariance described by a genomic relationship matrix. The model was implemented via the EMMAX software [6].

Based on their ID number, SNPs were annotated to genes corresponding to the ARS-UCD1.2 reference genome using the Bioconductor BioMart tool version 3.14 [7] and then genes were annotated to KEGG reference pathways (map) using the David software version 6.8 [8]. The effects of KEGG pathways on stature were estimated separately for each breed using the following mixed linear model that accounted for the similarity between pathways:

$$\mathbf{y} = \mathbf{1}\boldsymbol{\mu} + \mathbf{Z}\mathbf{t} + \mathbf{e},\tag{1}$$

where **y** is the vector of absolute values of SNP additive effects on stature that are estimated separately for each breed in the GWAS of Bouwman et al. [4], **1** is a vector of ones, μ represents the general mean, **t** is the vector of the random effects of KEGG pathways with a preimposed normal distribution defined by N(0, $V\sigma_t^2$), **e** is the vector of residuals distributed as N(0, $I\sigma_e^2$), **Z** is the incidence matrix for **t**. Note that if multiple SNPs were identified within a gene, only one SNP with the strongest effect was included in **y**, so that each gene is represented by a single variant. The similarity between KEGG pathways i and j, was introduced into the model by incorporating a non-diagonal KEGG covariance matrix **V**. This covariance was expressed by the Jaccard similarity coefficient:

$$J(i,j) = \frac{M}{N},$$
(2)

where M represents the number of genes shared between KEGG pathways i and j, while N represents the total number of genes involved in KEGG pathways i and

j. Variance components were assumed to be known: $\sigma_t^2 = 0.3\sigma_v^2$ and $\sigma_e^2 = 0.7\sigma_v^2$.

The mixed model equations [9] were used to obtain solutions for μ and **t**:

$$\begin{bmatrix} \widehat{\mu} \\ \widehat{\mathbf{t}} \end{bmatrix} = \begin{bmatrix} \mathbf{1}^{\mathrm{T}} \mathbf{R}^{-1} \mathbf{1} & \mathbf{1}^{\mathrm{T}} \mathbf{R}^{-1} \mathbf{Z} \\ \mathbf{Z}^{\mathrm{T}} \mathbf{R}^{-1} \mathbf{1} & \mathbf{Z}^{\mathrm{T}} \mathbf{R}^{-1} \mathbf{Z} + \mathbf{G}^{-1} \end{bmatrix}^{-1} \begin{bmatrix} \mathbf{1}^{\mathrm{T}} \mathbf{R}^{-1} \mathbf{y} \\ \mathbf{Z}^{\mathrm{T}} \mathbf{R}^{-1} \mathbf{y} \end{bmatrix},$$
(3)

where $\mathbf{R} = \mathbf{I}\widehat{\sigma}_{e}^{2}$ and $\mathbf{G} = \mathbf{V}\widehat{\sigma}_{t}^{2}$.

To maximise the computational performance of the estimation/prediction process, a custom Python program implementing the NumPy 1.19.5 library [10] was used. Since all calculations were carried out on a highperformance server, the NumPy library was also used to set the array indexing and order, which further improved the computing time compared to a native Python application. Each element of t was assessed for significance $(H_0: \hat{t}_i \leq 0 \text{ vs. } H_1: \hat{t}_i > 0)$ by calculating the probability of obtaining a more extreme value from the N(0, σ_t^2) density function. Since NumPy and SciPy application programming interfaces (API) are implemented with LAPACK and BLAS, which require Fortran memory layout, all input matrices were transformed to Fortran to avoid costly transposing. In comparison to a fixed matrix input, this approach results in a ten times faster estimation process.

Results and discussion

The effects of 179 KEGG pathways were estimated based on the effects of selected SNPs from a whole-genome sequence-based GWAS of Bouwman et al. [4], separately for three Nordic cattle breeds-Danish Holstein (DH with 366,877 SNPs), Danish Red Dairy Cattle (DR with 299,723 SNPs), and Finnish Red Dairy Cattle (FR with 396,224 SNPs) (Fig. 1). In DH and FR, the same pathway-D-amino acid metabolism (map00473) revealed a significant effect on stature with moderate P-values of 0.035 (FR) and 0.049 (DH). In DR, it also reached a borderline significance of 0.133. Depending on the breed, the effect of map00473 was estimated based on 78 SNPs in DH and FR, and 76 SNPs in DR (Fig. 2 and Additional file 1: Tables S1 and S2). The differences in SNP counts resulted from the fact that the input SNP panel in Bouwman et al. [4] was pre-processed separately for each breed, which resulted in breed-specific SNP exclusion. In addition, the pathway responsible for the metabolism of terpenoids and polyketides (map01059) was significant (P=0.041) in DH, while the synthesis and degradation of ketone bodies pathway (map00072) and the pathway of biosynthesis of various plant secondary metabolites (map00999) were significant in DR with P = 0.037 and P = 0.047, respectively.

Kotlarz et al. Genetics Selection Evolution (2022) 54:80

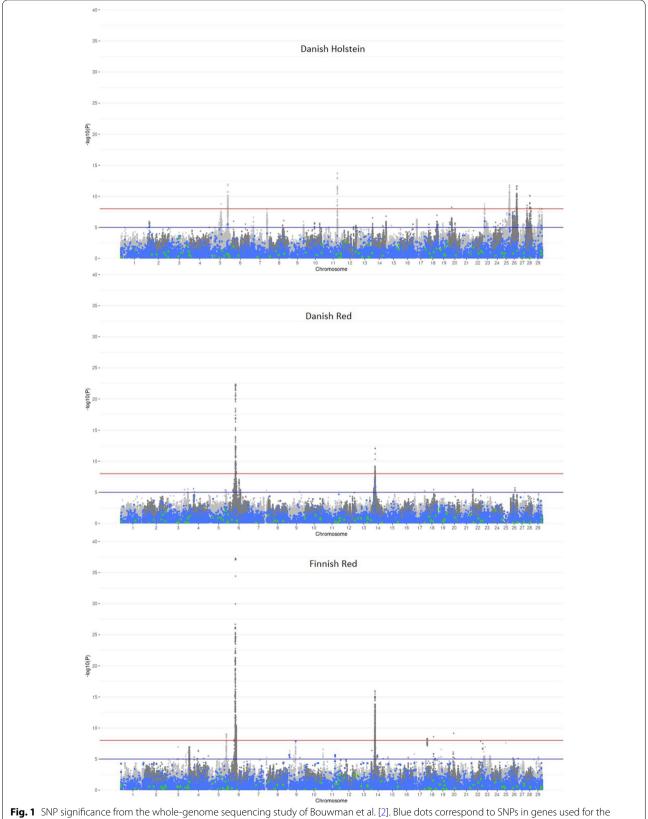
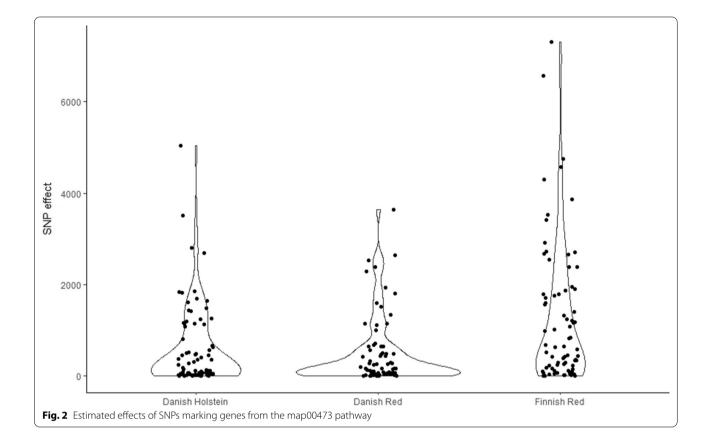


Fig. 1 SNP significance from the whole-genome sequencing study of Bouwman et al. [2]. Blue dots correspond to SNPs in genes used for the estimation of the effects of KEGG pathways in model (1), green dots correspond to SNPs marking genes constituting the map00473 pathway, and grey SNPs are the remainder



The idea to incorporate pathway effects directly into a mixed GWAS-based model of complex phenotypes dates back to 2014 and was presented by Evangelou et al. [11] who proposed two models and corresponding estimators of their effects. A difference between the models presented in [11] and our model is the use of measured phenotypes as a dependent variable in their case vs SNP summary statistics as a dependent variable in our case. Moreover, Evangelou et al. [11] defined no covariance between pathways, but allowed pathway-specific variances, while in our model we used a non-diagonal covariance.

When interpreting the effects of KEGG pathways, two scenarios emerge. On the one hand, the overall strong effect of a pathway may be driven by the strong effect of a single gene, that is this pathway's component—a situation that could have been detected in a conventional GWAS. On the other hand, the strong effect of a pathway may be due to the combined effects of many genes constituting this pathway—a situation that may easily be missed in GWAS due to the small or moderate effects of particular genes from the pathway. In the case of our data—none of the genes that harbour the most significant SNPs in the GWAS performed by Bouwman et al. [4] were a component of the D-amino acid metabolism pathway, which therefore leads to the conclusion that the whole pathway is a significant component of the genetic determination of stature. Biologically, an outstanding pattern of our study was that the pathway associated with the metabolism of D-amino acids is significant for two breeds and on the border of claimed significance in the third breed. Although D-amino acids do not occur in naturally-translated proteins, the link between D-amino acids metabolism and growth has long been recognised. Experimentally, a supplementation of mice with D-amino acids resulted in increased weight that was observed with an increased concentration of D-phenylalanine and D-tryptophan in the diet [12]. Moreover, D'Aniello [13] reported that, in the pituitary gland, D-aspartic acid stimulates the secretion of the growth hormone in rats. In cattle, a supplementation of food with synthetic amino acids is a very common practice with commercial diet supplements containing a mixture of naturally occurring L-versions as well as not naturally occurring D-versions. Campbell et al. [14] observed that D-amino acids are somewhat less efficiently metabolised than their naturally occurring synonyms. Since methionine is often the first limiting amino acid for growth in cattle [15], individuals that possess a more efficient mechanism of D-amino acid metabolism are expected to grow better, which may result in higher stature in adults.

Another metabolic pathway demonstrating potential importance on stature is the synthesis and degradation of the ketone bodies pathway (map00072) that was significant in DR. It has been demonstrated that the metabolism of ketone bodies is related to the growth of the whole organism (mainly through the SLC16A6 gene as reported by Kichaev et al. [16] and Karanth et al. [17]) and also that it acts at a single-cell level by triggering mitochondrial response towards cell's oxidative stress and deficiency in metabolic energy [18]. Although the other pathway of biosynthesis of various plant secondary metabolites that was significant in DR, does not relate directly to animal metabolism, it can be hypothesised that genes playing a role in the biochemical processing of metabolites originating from plants lead to higher feed efficiency in cattle and furthermore influence animals' growth, but experimental evidence is lacking.

Conclusions

Our results demonstrate that taking higher-order components of biological systems, such as metabolic pathways, into consideration, provides valuable insight into the basis of the variation of complex phenotypes, that may be missed by conventional GWAS and should be used as an enhancement thereof. From a practical perspective, it would imply conducting genomic selection not only on the most significant, major genes but also considering "additional" genes that are members of the significant metabolic pathways. This shall be especially useful for phenotypes that have undergone unidirectional selection for several decades, such as milk production traits, for which it can be expected that most of the major genes are already close to being homozygous for causal mutations.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12711-022-00771-1.

Additional file 1: Table S1. Gene effects in map00473. Estimated gene effects (represented by the most significant SNP) constituting the map00473 KEGG pathway. Table S2. KEGG pathway effects. Estimated effects of all KEGG pathways. Significant pathways ($P \le 0.05$) are marked in blue.

Acknowledgements

The genome-wide association studies were part of the Center for Genomic Selection in Animals and Plants (GenSAP) financed by Innovation Fund Denmark (Grant: 0603-00519B). The 1000 Bull Genomes Project is acknowledged for sharing sequence data for imputation. We thank prof. Bernt Guldbrandtsen for fruitful discussions on pathway modelling. Calculations have been

carried out using resources provided by Wroclaw Centre for Networking and Supercomputing.

Author contributions

KK performed all data analyses, BK acquired and edited the data, ZC and GS interpreted the data, JS designed the work, interpreted the data and drafted the manuscript. All authors contributed to the writing of the manuscript, reviewed and approved its final version. All authors read and approved the final manuscript.

Funding

Not applicable.

Availability of data and materials

BioSample accession codes to raw data are provided in https://doi.org/10. 1038/s41588-018-0056-5.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Biostatistics Group, Department of Genetics, The Wroclaw University of Environmental and Life Sciences, Kozuchowska 7, 51-631 Wroclaw, Poland. ²Center for Quantitative Genetics and Genomics, Aarhus University, Blichers Allé 20, 8830 Tjele, Denmark. ³National Research Institute of Animal Production, Krakowska 1, 32-083 Balice, Poland.

Received: 11 September 2022 Accepted: 6 December 2022 Published online: 16 December 2022

References

- Manolio TA, Collins FS, Cox NJ, Goldstein DB, Hindorff LA, Hunter DJ, et al. Finding the missing heritability of complex diseases. Nature. 2009;461:747–53.
- Khatri P, Sirota M, Butte AJ. Ten years of pathway analysis: current approaches and outstanding challenges. PLoS Comput Biol. 2012;8:e1002375.
- Stingo FC, Chen YA, Tadesse MG, Vannucci M. Incorporating biological information into linear models: a Bayesian approach to the selection of pathways and genes. Ann Appl Stat. 2011;5:1978–2002.
- Bouwman AC, Daetwyler HD, Chamberlain AJ, Ponce CH, Sargolzaei M, Schenkel FS, et al. Meta-analysis of genome-wide association studies for cattle stature identifies common genes that regulate body size in mammals. Nat Genet. 2018;50:362–7.
- Fuchsberger C, Abecasis GR, Hinds DA. minimac2: faster genotype imputation. Bioinformatics. 2015;31:782–4.
- Kang HM, Sul JH, Service SK, Zaitlen NA, Kong SY, Freimer NB, et al. Variance component model to account for sample structure in genomewide association studies. Nat Genet. 2010;42:348–54.
- Smedley D, Haider S, Ballester B, Holland R, London D, Thorisson G, Kasprzyk A. BioMart–biological queries made easy. BMC Genomics. 2009;10:22.
- Huang DW, Sherman BT, Tan Q, Collins JR, Alvord WG, Roayaei J, et al. The DAVID Gene Functional Classification Tool: a novel biological modulecentric algorithm to functionally analyze large gene lists. Genome Biol. 2007;8:R183.
- 9. Henderson CR. Applications of linear models in animal breeding. Guelph: University of Guelph Press; 1984.
- 10. Harris CR, Millman KJ, van der Walt SJ, Gommers R, Virtanen P, Cournapeau D, et al. Array programming with NumPy. Nature. 2020;585:357–62.

- 11. Evangelou M, Dudbridge F, Wernisch L. Two novel pathway analysis methods based on a hierarchical model. Bioinformatics. 2014;30:690–7.
- 12. Friedman M, Levin CE. Nutritional and medicinal aspects of D-amino acids. Amino Acids. 2012;42:1553–82.
- 13. D'Aniello A. D-Aspartic acid: an endogenous amino acid with an important neuroendocrine role. Brain Res Rev. 2006;53:215–34.
- 14. Campbell CG, Titgemeyer EC, St-Jean G. Efficiency of D- vs L-methionine utilization by growing steers. J Anim Sci. 1996;74:2482–7.
- Richardson CR, Hatfield EE. The limiting amino acids in growing cattle. J Anim Sci. 1978;46:740–5.
- Kichaev G, Bhatia G, Loh PR, Gazal S, Burch K, Freund MK, et al. Leveraging polygenic functional enrichment to improve GWAS power. Am J Hum Genet. 2019;104:65–75.
- Karanth S, Schlegel A. The monocarboxylate transporter SLC16A6 regulates adult length in zebrafish and is associated with height in humans. Front Physiol. 2019;9:1936.
- Kolb H, Kempf K, Röhling M, Lenzen-Schulte M, Schloot NC, Martin S. Ketone bodies: from enemy to friend and guardian angel. BMC Med. 2021;19:313.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

